Реагенты для экстракции белка

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Servicebio®Coomassie Brilliant Blue G250

Cat #: G2039-250ML

Product Information

Product Name	Cat. No.	Spec.
Coomassie Brilliant Blue G250	G2039-250ML	250 mL

Product Description/Introduction

Coomassie Brilliant Blue G250 can bind proteins rapidly. The maximum absorption of Coomassie Brilliant Blue G250 is 488 nm in the free state. After binding to protein, the maximum absorption changed to 595 nm. The optical absorption value is proportional to the protein content, so it can be used for the quantitative detection of protein. This product can also be used as a supplement to the G2001 Bradford Assay protein quantitative detection kit.

Storage and Shipping Conditions

Ship and store at room temperature, valid for 12 months.

Assay Protocol/Procedures

1. **(Optional) Drawing standard curve (microplate reader method):** Dissolve BSA in water to prepare 0.5 mg/mL protein standard working solution. The protein standard working solution is added to the 96-well plate at levels of 0, 1, 2, 4, 8, 12, 16, 20 μL, and then the gradient working solution is supplemented to 20 μL with PBS or saline. The gradient curves of protein concentration are 0, 25, 50, 100, 200, 300, 400 and 500 μg/mL. If only relative quantitative comparisons are made, no standard curves are required.
2. **Prepare the sample to be tested:** The protein sample to be tested is diluted appropriately (the protein concentration of the sample can be detected by pre-experiment to be within the range of the standard curve, to ensure the reliability of the test result), and 20 μL of each sample is added to the 96-well plate. The sample to be tested should be diluted in the same solution as the protein standard.
3. **Detection:** 200 μL Coomassie brilliant blue G250 solution is added to each well and thoroughly mixed (the 96-well plate can be placed on the oscillator for 30 s). After 3-5 min at room temperature, the standard curve No. 0 is used as a reference, and the colorimetric measurement is performed at 595 nm wavelength, and the absorbance value of each well is recorded.
4. **Calculation:** The gradient protein content (μg/mL) in the standard curve is taken as the abscissa, and the light absorption value is taken as the ordinate to draw the standard curve. According to the absorbance value of the sample, the protein concentration of the sample to be measured in the corresponding well can be found on the standard curve (μg/mL), and then multiplied by the dilution of the sample, the actual protein concentration of the sample to be measured.

   In addition: if the spectrophotometer is used to determine, the glass test tube or glass colorimetric tube is used as the reaction vessel to make standard curves. After proper dilution of the protein sample to be tested, add it to a new glass test tube or colorimetric tube with a sample size of 1 mL. Add 3 mL of Coomassie brilliant blue G250 solution to the standard curve gradient tube and sample tube, mix thoroughly, stand at room temperature for 3-5 min, then use spectrophotometer for colorimetric detection.

   Detection: The wavelength of the spectrophotometer is set to 595 nm, and the standard curve and the

sample to be tested are zeroed with the standard curve No. 0 tube as the reference. Standard curves are drawn as described in Step 4 and protein concentrations in the samples to be tested are calculated.

Note

1. The product should be restored to room temperature before use, and mixed upside down that avoid to affect the sensitivity of detection.
2. For your safety and health, please wear safety glasses, gloves, or protective clothing.

Servicebio® BCA Protein Quantitative Detection Kit

Cat #: G2026

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Product Information

Product Name	Cat. No.	Spec.
BCA Protein Quantitative Detection Kit	G2026-200T	200T
	G2026-1000T	1000T

Product Description/Introduction

Bradford and BCA methods are the most commonly used methods for quantitative determination of protein concentration. The BCA Protein Quantitative Detection Kit is a detergent-compatible formulation based on bicinchoninic acid (BCA) for the colorimetric detection and quantitation of total protein. This method combines the well-known reduction of Cu^{2+} to Cu^{+} by protein in an alkaline medium (the biuret reaction) with the highly sensitive and selective colorimetric detection of the cuprous cation (Cu^{+}) using a unique reagent containing bicinchoninic acid. The purple-colored reaction product of this assay is formed by the chelation of two molecules of BCA with one cuprous ion. This water-soluble complex exhibits a strong absorbance at 562 nm that is nearly linear with increasing protein concentrations over a broad working range (20–2000 μg/mL).

This method is not affected by chemical substances and high concentration of detergent in most samples, including 5% SDS, 5% Triton X-100 and 5% Tween-20, 60, 80, etc. However, chelating agent and high concentration reducing agent will affect the detection results. So, it is necessary to ensure that there is no EGTA in the sample, the concentration of EDTA is less than 10 mM, and the concentration of DTT and β-mercaptoethanol is less than 1mM. If the sample contains chelating agent or reducing agent, please consider Bradford protein quantitative detection kit (G2001).

Storage and Shipping Conditions

Ship at room temperature; Protein standard (BSA) store at 4℃, valid for 12 months. The protein standard solution should be stored at -20℃ and used within 6 months. The remaining reagents are stored at room temperature, valid for 12 months.

Product Components

Component Number	Component	G2026-200T	G2026-1000T
G2026-1	BCA reagent	40 mL	2×100 mL
G2026-2	Cupric sulfate solution	1.2 mL	5×1.2 mL
G2026-3	Protein standard tube (BSA)	25 mg	5×25 mg
G2026-4	Protein standard diluent solution	1.5 mL	5×1.5 mL
Manual		One copy	

Assay Protocol/Procedures

1. Preparation of protein standard storage solution: add 1 mL of protein standard diluent solution to protein standard tube (BSA), and 25 mg protein standard is completely dissolved to obtain protein standard storage solution with a concentration of 25 mg/mL. The standard protein storage solution can be stored for a long time at -20℃;
2. Preparation of protein standard working solution: 25 mg/ml protein standard storage solution is diluted 50 times with PBS or normal saline to obtain protein standard working solution with a final

concentration of 0.5 mg/ml. Pay attention to the 10 times gradient method for dilution to ensure accuracy.

3. Standard curve (Microplate Procedure): use the following table as a guide to prepare a set of protein standards:

Dilution Scheme for Microplate Procedure (Working Range = 0–500 μg/mL)			
96-well plates	Volume of Diluent (μL) (PBS or normal saline)	Volume of 0.5 mg/ml protein standard working solution (μL)	Final BSA Concentration (μg/mL)
A	20	0	0
B	19	1	25
C	18	2	50
D	16	4	100
E	12	8	200
F	8	12	300
G	4	16	400
H	0	20	500

4. Preparation of test-sample: the test-sample is diluted appropriately (through the pre-test detection, the protein concentration of the test-sample is within the range of the standard curve to ensure that the detection results are reliable), and pipette 20 μL of each test-sample into the 96-well plates. The test-sample and the protein standard working solution are diluted with the same solution.
5. Preparation of BCA working solution: mixing 50 parts of BCA Reagent with 1 part of copper sulfate solution (50:1, BCA Reagent: copper sulfate solution). BCA working solution can be stored at room temperature and used within 24h. Add 200 μL of the BCA working solution to each well. It is recommended to prepare as required to avoid waste.
6. Detection: add 200 μL of the BCA working solution to each well and mix plate thoroughly on a plate shaker for 30 seconds; After cover plate and incubate at 37°C for 30 minutes, the standard curve No. 0 was used as a reference to measure the absorbance at or near 562 nm on a plate reader. (Note: it can also be reacted at room temperature for 2 h or 60℃ for 30 min. If the protein concentration is low, it is recommended to react at 60℃)
7. Calculation: the standard curve was drawn with gradient protein content (μg/mL) as abscissa and absorption value as ordinate. According to the absorbance value of the test-sample, the protein concentration (μg/mL) of the test-sample in the corresponding well can be found on the standard curve, and then multiplied by the dilution factor of the sample is the actual protein concentration of the test-sample.

Note

1. Determination of protein concentration by BCA method is greatly affected by temperature and time, and the absorbance value will change with the extension of time or the increase of temperature. If the time and temperature of color reaction cannot be accurately controlled, it is recommended to make a standard curve for each determination.
2. When preparing protein standard storage solution, it is necessary to ensure sufficient dissolution. Dilution 10 times gradient dilution is recommended when preparing protein standard working solution. Do not dilute 50 times at a time to avoid error.
3. In order to guarantee quantification of protein, it is better to choose the same buffer solution for sample extraction and protein standard dilution to ensure the same detection conditions. If the buffer has a high background value, other methods are recommended.
4. For your safety and health, please wear safety glasses, gloves, or protective clothing.

Servicebio® BCA Protein Quantitative Detection Kit

Cat #: G2026

Product Information

Product Name	Cat. No.	Spec.
BCA Protein Quantitative Detection Kit	G2026-200T	200 T
	G2026-1000T	1000 T

Product Description/Introduction

Bradford and BCA methods are the most commonly used methods for quantitative determination of protein concentration. The BCA Protein Quantitative Detection Kit is a detergent-compatible formulation based on bicinchoninic acid (BCA) for the colorimetric detection and quantitation of total protein. This method combines the well-known reduction of Cu^{2+} to Cu^{+} by protein in an alkaline medium (the biuret reaction) with the highly sensitive and selective colorimetric detection of the cuprous cation (Cu^{+}) using a unique reagent containing bicinchoninic acid. The purple-colored reaction product of this assay is formed by the chelation of two molecules of BCA with one cuprous ion. This water-soluble complex exhibits a strong absorbance at 562 nm that is nearly linear with increasing protein concentrations over a broad working range (50–2000 μg/mL).

This method is not affected by chemical substances and high concentration of detergent in most samples, including 5% SDS, 5% Triton X-100 and 5% Tween-20, 60, 80, etc. However, chelating agent and high concentration reducing agent will affect the detection results. So, it is necessary to ensure that there is no EGTA in the sample, the concentration of EDTA is less than 10 mM, and the concentration of DTT and β-mercaptoethanol is less than 1mM. If the sample contains chelating agent or reducing agent, please consider Bradford protein quantitative detection kit (G2001).

Storage and Shipping Conditions

Ship at room temperature; Protein standard (BSA) store at 2-8℃, valid for 12 months. The protein standard solution should be stored at -20℃ and used within 6 months. The remaining reagents are stored at room temperature, valid for 12 months.

Product Components

Component Number	Component	G2026-200T	G2026-1000T
G2026-1	BCA reagent	40 mL	2×100 mL
G2026-2	Cupric sulfate solution	1.2 mL	5×1.2 mL
G2026-3	Protein standard tube (BSA)	25 mg	5×25 mg
G2026-4	Protein standard diluent solution	1.5 mL	5×1.5 mL
Manual		One copy	

Assay Protocol/Procedures

1. Preparation of protein standard storage solution: add 1 mL of protein standard diluent solution to protein standard tube (BSA), and 25 mg protein standard is completely dissolved to obtain protein standard storage solution with a concentration of 25 mg/mL. The standard protein storage solution can be stored for a long time at -20℃;
2. Preparation of protein standard working solution: 25 mg/ml protein standard storage solution is diluted 50 times with PBS or normal saline to obtain protein standard working solution with a final

concentration of 0.5 mg/ml. Pay attention to the 10 times gradient method for dilution to ensure accuracy.

3. Standard curve (Microplate Procedure): The protein standard working solution was added to the 96-well plate at 0, 1, 2, 4, 8, 12, 16 and 20 μL, and then the above gradient working solution was replenished to 20 μL by adding 20, 19, 18, 16, 12, 8, 4 and 0 μL with PBS or physiological saline in order to obtain the gradient curves with the concentrations of protein at 0, 25, 50, 100, 200, 300, 400 and 500 μg/mL in the following order. The gradient curves were obtained in the order of 0, 25, 50, 100, 200, 300, 400, 500 μg/mL.
4. Preparation of test-sample: the test-sample is diluted appropriately (through the pre-test detection, the protein concentration of the test-sample is within the range of the standard curve to ensure that the detection results are reliable), and pipette 20 μL of each test-sample into the 96-well plates. The test-sample and the protein standard working solution are diluted with the same solution.
5. Preparation of BCA working solution: mixing 50 parts of BCA Reagent with 1 part of copper sulfate solution (50:1, BCA Reagent: copper sulfate solution). BCA working solution can be stored at room temperature and used within 24 h. Add 200 μL of the BCA working solution to each well. It is recommended to prepare as required to avoid waste.
6. Detection: add 200 μL of the BCA working solution to each well and mix plate thoroughly on a plate shaker for 30 seconds; After cover plate and incubate at 37°C for 30 minutes, the standard curve No. 0 was used as a reference to measure the absorbance at or near 562 nm on a plate reader. (Note: it can also be reacted at room temperature for 2 h or 60℃ for 30 min. If the protein concentration is low, it is recommended to react at 60℃)
7. Calculation: the standard curve was drawn with gradient protein content (μg/mL) as abscissa and absorption value as ordinate. According to the absorbance value of the test-sample, the protein concentration (μg/mL) of the test-sample in the corresponding well can be found on the standard curve, and then multiplied by the dilution factor of the sample is the actual protein concentration of the test-sample.

Note

1. Determination of protein concentration by BCA method is greatly affected by temperature and time, and the absorbance value will change with the extension of time or the increase of temperature. If the time and temperature of color reaction cannot be accurately controlled, it is recommended to make a standard curve for each determination.
2. When preparing protein standard storage solution, it is necessary to ensure sufficient dissolution. Dilution 10 times gradient dilution is recommended when preparing protein standard working solution. Do not dilute 50 times at a time to avoid error.
3. In order to guarantee quantification of protein, it is better to choose the same buffer solution for sample extraction and protein standard dilution to ensure the same detection conditions. If the buffer has a high background value, other methods are recommended.
4. For your safety and health, please wear safety glasses, gloves, or protective clothing.

Servicebio® Coomassie (Bradford) Protein Quantitative Assay Kit

Cat #: G2001-250ML

Product Information

Product Name	Cat. No.	Spec.
Coomassie (Bradford) Protein Quantitative Assay Kit	G2001-250ML	250 mL

Product Description/Introduction

Two of the most commonly used methods for quantifying protein concentration are the Bradford method and the BCA method. The principle of the Bradford method is based on the rapid binding of the Coomassie brilliant blue G250 to proteins. The maximum absorption of Coomassie Brilliant Blue G250 is 488 nm in the free state. After binding to protein, the maximum absorption changed to 595 nm. and the light absorption value is proportional to the content of proteins, so it can be used for the quantitative detection of proteins. This method determines protein concentrations independent of the chemicals in the vast majority of samples, which can be as high as 1 mM for β-mercaptoethanol and 5 mM for dithiothreitol; However, affected by slightly higher concentrations of descaling agents, it is necessary to ensure that the SDS content is less than 0.01%, the Triton X-100 content is less than 0.05%, and the Tween-20, Tween-60, Tween-80, etc. content is less than 0.015%. The BCA Protein Quantitative Detection Kit (G2026) manufactured by our company is recommended for samples containing descaling agents.

This kit is easy and fast to operate, with high sensitivity and extremely fast detection speed, and can detect protein concentrations ranging from 25-1000 μg/mL.

Storage and Shipping Conditions

Product shipped at room temperature. Bovine Serum Albumin (BSA) Standard Ampules stored at 4℃, valid for 12 months. The protein standard solution, prepared with BSA Standard Ampules and Standard Ampules Dilution, store at -20°C and used within 6 months.

Product Components

Component Number	Component	G2001-250ML
G2001-1	Coomassie G-250 dye	250 mL
G2001-2	Bovine Serum Albumin (BSA) Standard Ampules	25 mg
G2001-3	Standard Ampules Dilution	1.5 mL
Manual		1 pc

Assay Protocol/Procedures

1. Preparation of **Protein Standard Stock Solution:** Dilute 25 mg Bovine Serum Albumin (BSA) Standard Ampules into 1 mL Standard Ampules Dilution; The concentration of **Protein standard stock solution** is 25 mg/mL, and it should be stored at -20℃.
2. Preparation of Protein Standard Working Solution: An appropriate amount of 25 mg/mL **Protein Standard Stock Solution** is diluted 50 times with PBS or normal saline to obtain a **Protein Standard Working Solution** with a final concentration of 0.5 mg/mL. Take care to dilute according to the 10-fold gradient method to ensure accurate dilution.
3. Preparation of the standard curve (microplate reader method): The protein standard working solution is added to 96-well plate at 0, 1, 2, 4, 8, 12, 16, 20 μL, and then PBS or normal saline is added to the same 96-well plate in turn at 20, 19, 18, 16, 12, 8, 4, 0 μL to make up the above gradient working

solution to 20 μL. The protein concentrations of Gradient curves is 0, 25, 50, 100, 200, 300, 400, and 500 μg/mL.

4. Preparation of the sample: Dilute the sample to be tested appropriately (by pre-experimental detection, the protein concentration of the sample can be within the range of the standard curve to ensure the reliability of the test result). Add 20 μL of samples to a 96-well plate. The sample to be tested and the protein standard are diluted with the same solution.
5. Test Tube Protocol:
   a) Add 200 μL of the Coomassie G-250 dye to each tube and mix well (The 96-well plate can be shaken on a vortex for 30 s).
   b) Incubate samples for 3-5 minutes at room temperature (RT).
   c) The standard curve No. 0 was used as a reference and colorimetrically measured at 595 nm, and the absorbance value of each well was recorded.
6. Calculation: Take the gradient protein content (μg/mL) in the standard curve as the horizontal coordinate and the absorbance value as the vertical coordinate, and plot the standard curve. According to the absorbance value of the measured sample, the protein concentration (μg/mL) of the sample to be tested in the corresponding wells can be found on the standard curve, and then multiplied by the dilution of the sample that is the actual protein concentration of the sample to be tested.

   Also: if measured by spectrophotometer, change the gradient standard reaction volume to 1 mL in step 3. Add 0, 0.05, 0.1, 0.2, 0.4, 0.6, 0.8, 1.0 mL of protein standard working solution (concentration of 0.5 mg/mL) to 7 clean glass test tubes or colorimetric tubes, then add 1.0, 0.95, 0.9, 0.8, 0.6, 0.4, 0.2, 0 mL of PBS or saline to each tube in order, and then top up the gradient working solution to 1 mL. The working solution of the gradient was replenished to 1 mL, and each tube was mixed thoroughly to obtain the gradient curve with protein concentrations of 0, 25, 50, 100, 200, 300, 400, 500 μg/mL.

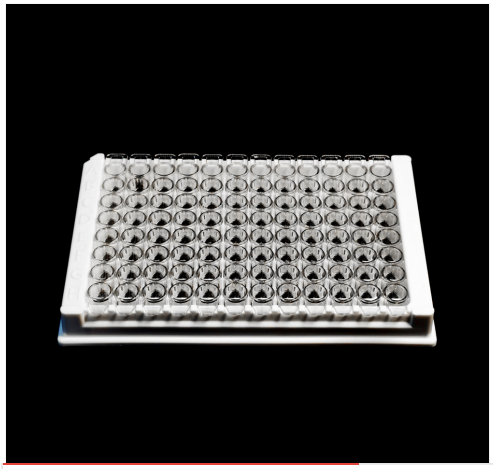
   Protein samples to be tested were made appropriately diluted and added to new glass test tubes or colorimetric tubes with a sample volume of 1 mL.

   Add 3 mL of Coomassie G-250 dye to each of the above standard curve gradient tubes and sample tubes, mix thoroughly, and let it stand for 3-5 min at room temperature for colorimetric detection by spectrophotometer.

   The wavelength of the photometer was set at 595 nm during the detection, and the standard curve and the sample to be tested were detected with the standard curve No. 0 tube as the reference for zero adjustment. Plot the standard curve and calculate the protein concentration in the sample to be tested according to the method in step 6.

Note

1. Coomassie G-250 dye should be brought to room temperature and mixed upside down before use to avoid affecting the sensitivity of the assay.
2. When preparing the protein standard stock solution, make sure that it is well dissolved. It is recommended to dilute the protein standard working solution in a 10-fold gradient, do not dilute 50 times at a time to avoid large errors.
3. Please wear safety glasses, gloves, or protective clothing.



Elisa Plate

Cat.No. : ESP-96-D

Brand : Servicebio

Spec.: 8-strip Well, Flat Bottom(10 pcs/bag) 200pcs/box

Product Introduction

Product Information

Cat.No.	Model	Specification	Material
ESP-96-D	8-stripWell, Flat Bottom	10 pcs/bag，200 pcs/carton	PS

Introduction

1. It is made of highly transparent polystyrene material with special surface treatment process, which canachieve high protein adsorption effect (> 600 ng /cm²); High sensitivity, saving coated protein.
2. Good stability,The coefficient of variation (CV) between wells is less than 5%;
3. Uniform and flat bottom design, light transmittance reach 90%
4. The bottom is even and flat, low refractive index, high transmittance, it is helpful to read the signal from bottom.
5. 8-strip well, 96Test, detachable, smart and economic
6. The maximum volume for each hole is around350μL.

Product Information

1、Bottom: Flat Bottom

2、Color:Transparent

3、Material:PS

4、Surface treatment: special treatment for high protein adsorption

5、Package: 10 pcs/bag，200 pcs/carton

6、Size (bag):14*9*15.5 cm

7、Size (Carton):47.5*29.5*33 cm

Storageand Transportation

Room temperature for storage andtransportation valid time: 24 months

Note

1.The item is not essential to have ‘national medical equipment production license’;

2.Please wear clothinglaboratory and disposable gloves when do operation.

Servicebio® PMSF (100mM)

Cat #: G2008-1ML

Product Information

Product Name	Cat. No.	Spec.
PMSF (100mM)	G2008-1ML	1 mL

Product Description/Introduction

The synthesis and breakdown of proteins in cells are in dynamic balance under stable conditions. However, when extracting proteins from cells and tissues in vitro, cells contain many endogenous proteases that can degrade proteins, such as phosphatases, serine proteases, and so on. These ubiquitous endogenous proteases can lead to the decomposition of target proteins, thus affecting the experimental results. Protease inhibitors are used to protect proteins from degradation by endogenous proteases released during protein extraction. PMSF (Phenylmethanesulfonyl fluoride, also know as phenylmethylsulfonyl fluoride, molecular formula $C_7H_7FO_2S$, molecular weight 174.19), can irreversibly inhibit serine protease (such as trypsin, chymotrypsin, thrombin) and thiol protease (such as papain).

Storage and Shipping Conditions

Ship with wet ice; Store at -20℃, valid for 12 months.

Product Components

Component	G2008-1ML
PMSF (100 mM)	1 mL
Manual	

Assay Protocol/Procedures

1. The concentration of this product is PMSF 100 mM, and the final concentration of working solution is recommended to be 1 mM.
2. This product is added to protein lysate at a ratio of 1:100 about 5 minutes before use, and mix well for reserve use.

Note

1. PMSF is highly toxic and seriously damages the mucosa of the respiratory tract, eyes and skin. It is fatal if inhaled, swallowed or absorbed through the skin. If PMSF comes into contact with eyes or skin, rinse immediately with plenty of water.
2. PMSF is unstable in aqueous solution and should be added to lysis buffer from storage solution before use.
3. For your safety and health, please wear safety glasses, gloves, or protective clothing.

Servicebio® Phosphoprotease Inhibitor

Cat #: G2007-1ML

Product Information

Product Name	Cat. No.	Spec.
Phosphoprotease Inhibitor	G2007-1ML	2×1 mL

Product Description/Introduction

The synthesis and breakdown of proteins in cells are in dynamic balance under stable conditions. However, when extracting proteins from cells and tissues in vitro , cells contain many endogenous proteases that can degrade proteins, such as phosphatases and serine proteases. These ubiquitous endogenous proteases can lead to the decomposition of target proteins, thus affecting the experimental results. Protease inhibitors are used to protect proteins from degradation by endogenous proteases released during protein extraction. Phosphatase inhibitors are also essential in the study of phosphorylated proteins.

The main components of this product are NaF and Na_3VO_4, which can inhibit tyrosine phosphatase, acid phosphatase and alkaline phosphatase.

Storage and Shipping Conditions

Ship with dry ice； Store at -20℃, valid for 12 months.

Product Components

Component Number	Component	G2007-1ML
G2007-1	Phosphoprotease inhibitor A solution (1 M NaF)	1 mL
G2007-2	Phosphoprotease inhibitor B solution (100 mM Na_3VO_4)	1 mL
Manual		

Assay Protocol/Procedures

1. Thaw and rewarm the product before use.
2. Mix well 1 μL of phosphoprotease inhibitor A solution and 1 μL of phosphoprotease inhibitor B solution to 100 μL of lysate, .

Note

1. Do not reuse if the product color turns yellow.
2. For your safety and health, please wear safety glasses, gloves, or protective clothing.

Servicebio® 50×Cocktail Protease Inhibitor

Cat #: G2006-250UL

Product Information

Product Name	Cat. No.	Spec.
50×Cocktail Protease Inhibitor	G2006-250UL	250 μL

Product Description/Introduction

Many of the cellular proteins are kept separate from proteolytic enzymes. Disruption of cellular and tissue architecture during protein extraction distorts the in vivo state by making all proteins potentially accessible for degradation or modification by endogenous proteases. That resulting unregulated proteolytic activity can reduce protein yield and function. Protease inhibitors can be added to the lysis reagents in order to prevent degradation of extracted proteins, and to obtain the best possible protein yield and activity following cell lysis.

50 × Cocktail Protease Inhibitor (EDTA-free) is a concentrated stock solution containing several different proteases inhibitor compounds (AEBSF, Aprotinin, Bestatin, E-64, Leupeptin, Pepstatin A) that can be added to samples to prevent protein degradation during cell lysis and protein extraction.

Storage and Shipping Conditions

Ship with dry ice. Store at -20°C, valid for 3 months.

Product Components

Component	G2006-250UL
50×Cocktail Protease Inhibitor	250μL
Manual	One copy

Assay Protocol/Procedures

This product is 50× concentrated solution. 20 μL of concentrated solution is added to 1 mL of protein lysis solution to ensure complete protection for protein extracts. It must be added before use and cannot be stored to prevent invalidation. At the same time, we provide protein lysis reagents (G2002, G2033).

Note

For your safety and health, please wear safety glasses, gloves, or protective clothing.

Servicebio® IP Lysis Buffer

Cat #: G2038-100ML

Product Information

Product Name	Cat. No.	Spec.
IP Lysis Buffer	G2038-100ML	100 mL

Product Description/Introduction

IP Lysis Buffer is a lysate that lysates cells or tissues to prepare protein samples under non-denaturing conditions. Protein samples obtained from tissue or cell lysis with this lysate can be applied to PAGE, western blot and immunoprecipitation (IP), co-immunoprecipitation (co-IP), Chromatin Immunopre-cipitation (ChIP) and ELISA where protein activity is required.

This product consists of 25 mM Tris-HCl, 150 mM NaCl, 1 mM EDTA and 1% NP-40 can be applied to animal or plant tissue and cell samples,as well as for fungal or bacterial samples.

Storage and Shipping Conditions

Ship with wet ice； Store away from light at 2-8℃, valid for 12 months.

Product Components

Component	G2038-10ML
IP Lysis Buffer	100 mL
Manual	

Assay Protocol/Procedures

Requires self-provide protease inhibitors. Protease inhibitors (G2006, G2007, G2008, etc.) should be added to IP lysate before use to prevent protein degradation. All references to IP lysates in the following instructions have been added with protease inhibitors.

For tissue samples:

1. The tissue blocks are washed with pre-cooled PBS (G4202 recommended) to remove blood stains, and then cut into small pieces and placed in the homogenizer.
2. Add 10 times tissue volume of IP Lysis Buffer and homogenize at low temperature (It is recommended to use high-speed tissue grinding instrument KZ-III-F and KZ-III-FP, which is independently developed and produced by Servicebio). Note that the amount of IP Lysis Buffer can be added at a ratio of approximately 50 mg of tissue to 1 mL of lysate. If the tissue protein content is low, the amount of lysate can be reduced to increase the protein concentration in the crude extract solution.
3. Transfer the homogenate to a 1.5mL of centrifuge tube and shake. Ice bath for 30 min with repeated pipetting every 10 min to ensure complete lysis of tissue cells;
4. Centrifuge at 12000 x g for 5 min and collect the supernatant, as total protein solution.

For adherent cell samples:

1. Wash the cells with PBS 2-3 times, and aspirate the residual liquid thoroughly at the last time.
2. Absorb IP Lysis Buffer into the cell culture plate and flask at the ratio of 250 μL of lysate per well of the 6-well plate, shake the culture plate and flask repeatedly to make the lysate fully contact with the cells for 3-5 min.
3. Scrape off the cells with a cell scraper and collect them into a centrifuge tube.

4. Centrifuge at 12000 x g for 5 min and collect the supernatant as total protein solution.

For suspended cell samples:

1. Cells were collected by centrifugation.
2. Mix the cytosol with IP lysate at a ratio of 250 μL of lysate per cell well of a 6-well plate,and shake.
3. Ice bath for 30 min, repeatedly pipetting every 10 min to ensure complete cell lysis.
4. Centrifuge at 12000 x g for 5 min and collect the supernatant as total protein solution.

For bacterial or fungal samples:

1. 1 mL of the bacterial suspension was centrifuged to remove the supernatant and washed once with PBS to fully remove the liquid. The vortices make the thalli disperse as much as possible.
2. Add 100-200 μL of IP lysate and vortex gently to mix well with the lysate..
3. Ice bath for 10 min, repeatedly pipetting every 2 min to ensure complete lysis of thalli.
4. 4. Centrifuge at 12000 x g for 5 min and collect the supernatant as total protein solution.

Note

1. Tissue or cells may appear viscous when lysed. This can be achieved by repeatedly blowing with a pipettor or shaking with a vortexer until it becomes a liquid. Add a further appropriate amount of lysis solution if it remains thick.
2. This reagent does not contain protease inhibitors, Add your own protease inhibitors before use. G2006, G2007, G2008 and other related protease inhibitors of our company are recommended.
3. The total protein solution obtained by cleavage of this product is compatible with the BCA protein quantitative detection kit (G2026) of our company.
4. For your safety and health, please wear safety glasses, gloves, or protective clothing.

Servicebio® RIPA Lysis Buffer (Weak)

Cat #: G2033

Product Information

Product Name	Cat. No.	Spec.
RIPA Lysis Buffer (Weak)	G2033-30ML	30 mL
	G2033-100ML	100 mL

Product Description/Introduction

RIPA Lysis Buffer is a traditional rapid lysis solution for cells and tissues. It is available in a variety of formulations and classified as strong, medium or weak. The protein samples obtained from the lysis of tissues and cells in RIPA weak lysis solutions can be used for routine PAGE, Western and other experiments where protein activity is not strictly required. This product contains 50 mM Tris-HCl (pH 7.4), 150 mM NaCl, 1 mM EDTA-2Na, 0.25% sodium deoxycholate and 1% NP-40. It is suitable for animal or plant tissues and cells, but can also be used for fungal or bacterial samples.

Storage and Shipping Conditions

Ship with wet ice; Store at 2-8℃ away from light; Valid for 12 months.

Product Components

Component	G2033-30ML	G2033-100ML
RIPA Lysis Buffer (Weak)	30 mL	100 mL
Manual		1 pc

Assay Protocol/Procedures

Requires self-provide protease inhibitors. Protease inhibitors (G2006, G2007, G2008, etc.) should be added to RIPA Lysis (Weak) before use to prevent protein degradation. All references to RIPA Lysate (Weak) in the following instructions have been added with protease inhibitors.

- **For tissue samples:**

1. The tissue pieces are washed with pre-cooled PBS (G4202 recommended) to remove blood stains, cut into small pieces and placed in the homogenizer.
2. Add 10 times the tissue volume of RIPA Lysis Buffer (Weak) and homogenate at low temperature (It is recommended to use high-speed tissue grinding instrument KZ-III-F and KZ-III-FP, which is independently developed and produced by Servicebio). Note that the amount of RIPA Lysis Buffer (Weak) can be added in a ratio of approximately 50 mg of tissue to 1 mL of lysate. If the content of tissue protein is low, the amount of lysate can be reduced to increase the protein concentration in crude extract solution.
3. Transfer the homogenate to a 1.5 mL of centrifuge tube and oscillate. Ice bath for 30 min with repeated pipetting every 10 min to ensure the complete lysis of tissue cells.
4. Centrifuge at 12000 x g for 5 min and collect the supernatant as total protein solution.

- **For adherent cell samples:**

1. Wash the cells with PBS for 2-3 times and aspirate the residual liquid thoroughly at the last time.
2. Aspirate RIPA Lysis Buffer (Weak) into the cell culture plate and flask at a ratio of 250 μL of cell lysate per well of 6-well plate, shake the culture plate and flask repeatedly to make the lysate fully contact with cells for 3-5 min.

3. Scrape off the cells with a cell scraper and collect into a centrifuge tube.
4. Lysis on ice for 30 min.
5. Centrifuge at 12000 x g for 5 min and collect the supernatant as total protein solution.

- **For suspension cell samples:**

1. Centrifuge to collect cells.
2. Mix the cytosol with RIPA Lysis Buffer (Weak) at a ratio of 250 μL of lysate per cell well of a 6-well plate and shake.
3. Ice bath for 30 min, repeatedly pipetting every 10 min to ensure complete cell lysis.
4. Centrifuge at 12000 x g for 5 min and collect the supernatant as total protein solution.

- **For bacterial or fungal samples:**

1. Take 1 mL of the suspension, centrifuge to remove the supernatant and wash once with PBS to fully remove the liquid. Vortex to disperse the bacterium as much as possible.
2. Add 100-200 μL of RIPA Lysis Buffer (Weak) and vortex gently to mix well with the lysate.
3. Ice bath for 30 min, repeatedly pipetting every 2 min to ensure complete cell lysis.
4. Centrifuge at 12000 x g for 5 min and collect the supernatant as total protein solution.

Note

1. Tissue or cells may appear viscous when lysed. This can be achieved by repeatedly blowing with a pipettor or shaking with a vortexer until it is liquid. Add a further appropriate amount of lysis solution if it remains thick.
2. This reagent does not contain protease inhibitors. Add your own protease inhibitors before use. G2006, G2007, G2008 and other related protease inhibitors of our company are recommended.
3. For your safety and health, please wear safety glasses, gloves, or protective clothing.

Servicebio® RIPA Lysis Buffer (Weak)

Cat #: G2033

Product Information

Product Name	Cat. No.	Spec.
RIPA Lysis Buffer (Weak)	G2033-30ML	30 mL
	G2033-100ML	100 mL

Product Description/Introduction

RIPA Lysis Buffer is a traditional rapid lysis solution for cells and tissues. It is available in a variety of formulations and classified as strong, medium or weak. The protein samples obtained from the lysis of tissues and cells in RIPA weak lysis solutions can be used for routine PAGE, Western and other experiments where protein activity is not strictly required. This product contains 50 mM Tris-HCl (pH 7.4), 150 mM NaCl, 1 mM EDTA-2Na, 0.25% sodium deoxycholate and 1% NP-40. It is suitable for animal or plant tissues and cells, but can also be used for fungal or bacterial samples.

Storage and Shipping Conditions

Ship with wet ice; Store at 2-8℃ away from light; Valid for 12 months.

Product Components

Component	G2033-30ML	G2033-100ML
RIPA Lysis Buffer (Weak)	30 mL	100 mL
Manual		1 pc

Assay Protocol/Procedures

Requires self-provide protease inhibitors. Protease inhibitors (G2006, G2007, G2008, etc.) should be added to RIPA Lysis (Weak) before use to prevent protein degradation. All references to RIPA Lysate (Weak) in the following instructions have been added with protease inhibitors.

- **For tissue samples:**

1. The tissue pieces are washed with pre-cooled PBS (G4202 recommended) to remove blood stains, cut into small pieces and placed in the homogenizer.
2. Add 10 times the tissue volume of RIPA Lysis Buffer (Weak) and homogenate at low temperature (It is recommended to use high-speed tissue grinding instrument KZ-III-F and KZ-III-FP, which is independently developed and produced by Servicebio). Note that the amount of RIPA Lysis Buffer (Weak) can be added in a ratio of approximately 50 mg of tissue to 1 mL of lysate. If the content of tissue protein is low, the amount of lysate can be reduced to increase the protein concentration in crude extract solution.
3. Transfer the homogenate to a 1.5 mL of centrifuge tube and oscillate. Ice bath for 30 min with repeated pipetting every 10 min to ensure the complete lysis of tissue cells.
4. Centrifuge at 12000 x g for 5 min and collect the supernatant as total protein solution.

- **For adherent cell samples:**

1. Wash the cells with PBS for 2-3 times and aspirate the residual liquid thoroughly at the last time.
2. Aspirate RIPA Lysis Buffer (Weak) into the cell culture plate and flask at a ratio of 250 μL of cell lysate per well of 6-well plate, shake the culture plate and flask repeatedly to make the lysate fully contact with cells for 3-5 min.

3. Scrape off the cells with a cell scraper and collect into a centrifuge tube.
4. Lysis on ice for 30 min.
5. Centrifuge at 12000 x g for 5 min and collect the supernatant as total protein solution.

- **For suspension cell samples:**

1. Centrifuge to collect cells.
2. Mix the cytosol with RIPA Lysis Buffer (Weak) at a ratio of 250 μL of lysate per cell well of a 6-well plate and shake.
3. Ice bath for 30 min, repeatedly pipetting every 10 min to ensure complete cell lysis.
4. Centrifuge at 12000 x g for 5 min and collect the supernatant as total protein solution.

- **For bacterial or fungal samples:**

1. Take 1 mL of the suspension, centrifuge to remove the supernatant and wash once with PBS to fully remove the liquid. Vortex to disperse the bacterium as much as possible.
2. Add 100-200 μL of RIPA Lysis Buffer (Weak) and vortex gently to mix well with the lysate.
3. Ice bath for 30 min, repeatedly pipetting every 2 min to ensure complete cell lysis.
4. Centrifuge at 12000 x g for 5 min and collect the supernatant as total protein solution.

Note

1. Tissue or cells may appear viscous when lysed. This can be achieved by repeatedly blowing with a pipettor or shaking with a vortexer until it is liquid. Add a further appropriate amount of lysis solution if it remains thick.
2. This reagent does not contain protease inhibitors. Add your own protease inhibitors before use. G2006, G2007, G2008 and other related protease inhibitors of our company are recommended.
3. For your safety and health, please wear safety glasses, gloves, or protective clothing.

Servicebio® RIPA Lysis Buffer (Strong)

Cat #: G2002

Product Information

Product Name	Cat. No.	Spec.
RIPA Lysis Buffer (Strong)	G2002-30ML	30 mL
	G2002-100ML	100 mL

Product Description/Introduction

RIPA Lysis Buffer is a traditional rapid lysis solution for cells and tissues. It is available in a variety of formulations and classified as strong, medium or weak. The protein samples obtained from the lysis of tissues and cells in RIPA strong lysis solutions can be used for routine PAGE, Western and other experiments where protein activity is not strictly required. This product contains 50 mM Tris-HCl (pH 7.4), 150 mM NaCl, 1 mM EDTA-2Na, 1% Triton X-100, 1% sodium deoxycholate and 0.1% SDS. It is suitable for animal or plant tissues and cells, but can also be used for fungal or bacterial samples.

Storage and Shipping Conditions

Ship with wet ice; Store in the dark at 2-8℃; valid for 18 months.

Product Components

Component	G2002-30ML	G2002-100ML
RIPA Lysis Buffer (Strong)	30 mL	100 mL
Manual	1 pc	

Preparation for The Experiment

Requires self-provide protease inhibitors. Protease inhibitors (G2006, G2007, G2008, etc.) should be added to RIPA Lysis (Strong) before use to prevent protein degradation. All references to RIPA Lysate (Strong) in the following instructions have been added with protease inhibitors.

Assay Protocol/Procedures

- **For tissue samples**

1. The tissue pieces are washed with pre-cooled PBS (G4202 recommended) to remove blood stains, cut into small pieces and placed in the homogenizer.
2. Add 10 times the tissue volume of RIPA lysis buffer (strong) and homogenate at low temperature (It is recommended to use high-speed tissue grinding instrument KZ-III-F and KZ-III-FP, which is independently developed and produced by Servicebio). Note that the amount of RIPA lysate (strong) can be added in a ratio of approximately 50 mg of tissue to 1 mL of lysate. If the content of tissue protein is low, the amount of lysate can be reduced to increase the protein concentration in crude extract solution.
3. Transfer the homogenate to a 1.5 mL of centrifuge tube and oscillate. Ice bath for 30 min with repeated pipetting every 10 min to ensure the complete lysis of tissue cells.
4. Centrifuge at 12000 x g for 5 min and collect the supernatant as total protein solution.

- **For adherent cell samples**

1. Wash the cells with PBS for 2-3 times and aspirate the residual liquid thoroughly at the last time.
2. Aspirate RIPA Lysis Buffer (Strong) into the cell culture plate and flask at a ratio of 250 μL of cell lysate

per well of 6-well plate, shake the culture plate and flask repeatedly to make the lysate fully contact with cells for 3-5 min.

3. Scrape off the cells with a cell scraper and collect into a centrifuge tube.
4. Lysis on ice for 30 min.
5. Centrifuge at 12000 x g for 5 min and collect the supernatant as total protein solution.

- **For suspension cell samples**

1. Centrifuge.to collect cells.
2. Mix the cytosol with RIPA Lysis Buffer (Strong) at a ratio of 250 μL of lysate per cell well of a 6-well plate and shake.
3. Ice bath for 30 min, repeatedly pipetting every 10 min to ensure complete cell lysis.
4. Centrifuge at 12000 x g for 5 min and collect the supernatant as total protein solution.

- **For bacterial or fungal samples**

1. Remove 1 mL of the suspension, centrifuge to remove the supernatant and wash once with PBS to fully remove the liquid. Vortex to disperse the bacterium as much as possible.
2. Add 100-200 μL of RIPA Lysis Buffer (Strong) and vortex gently to mix well with the lysate.
3. Ice bath for 30 min, repeatedly pipetting every 2 min to ensure complete cell lysis.
4. Centrifuge at 12000 x g for 5 min and collect the supernatant as total protein solution.

Note

1. Tissue or cells may appear viscous when lysed. This can be achieved by repeatedly blowing with a pipettor or shaking with a vortexer until it becomes a liquid. Add a further appropriate amount of lysis solution if it remains thick.
2. This reagent does not contain protease inhibitors. Add your own protease inhibitors before use. G2006, G2007, G2008 and other related protease inhibitors of our company are recommended.
3. For your safety and health, please wear safety glasses, gloves, or protective clothing.

Servicebio® RIPA Lysis Buffer (Strong)

Cat #: G2002

Product Information

Product Name	Cat. No.	Spec.
RIPA Lysis Buffer (Strong)	G2002-30ML	30 mL
	G2002-100ML	100 mL

Product Description/Introduction

RIPA Lysis Buffer is a traditional rapid lysis solution for cells and tissues. It is available in a variety of formulations and classified as strong, medium or weak. The protein samples obtained from the lysis of tissues and cells in RIPA strong lysis solutions can be used for routine PAGE, Western and other experiments where protein activity is not strictly required. This product contains 50 mM Tris-HCl (pH 7.4), 150 mM NaCl, 1 mM EDTA-2Na, 1% Triton X-100, 1% sodium deoxycholate and 0.1% SDS. It is suitable for animal or plant tissues and cells, but can also be used for fungal or bacterial samples.

Storage and Shipping Conditions

Ship with wet ice; Store in the dark at 2-8℃; valid for 18 months.

Product Components

Component	G2002-30ML	G2002-100ML
RIPA Lysis Buffer (Strong)	30 mL	100 mL
Manual	1 pc	

Preparation for The Experiment

Requires self-provide protease inhibitors. Protease inhibitors (G2006, G2007, G2008, etc.) should be added to RIPA Lysis (Strong) before use to prevent protein degradation. All references to RIPA Lysate (Strong) in the following instructions have been added with protease inhibitors.

Assay Protocol/Procedures

- **For tissue samples**

1. The tissue pieces are washed with pre-cooled PBS (G4202 recommended) to remove blood stains, cut into small pieces and placed in the homogenizer.
2. Add 10 times the tissue volume of RIPA lysis buffer (strong) and homogenate at low temperature (It is recommended to use high-speed tissue grinding instrument KZ-III-F and KZ-III-FP, which is independently developed and produced by Servicebio). Note that the amount of RIPA lysate (strong) can be added in a ratio of approximately 50 mg of tissue to 1 mL of lysate. If the content of tissue protein is low, the amount of lysate can be reduced to increase the protein concentration in crude extract solution.
3. Transfer the homogenate to a 1.5 mL of centrifuge tube and oscillate. Ice bath for 30 min with repeated pipetting every 10 min to ensure the complete lysis of tissue cells.
4. Centrifuge at 12000 x g for 5 min and collect the supernatant as total protein solution.

- **For adherent cell samples**

1. Wash the cells with PBS for 2-3 times and aspirate the residual liquid thoroughly at the last time.
2. Aspirate RIPA Lysis Buffer (Strong) into the cell culture plate and flask at a ratio of 250 μL of cell lysate

per well of 6-well plate, shake the culture plate and flask repeatedly to make the lysate fully contact with cells for 3-5 min.

3. Scrape off the cells with a cell scraper and collect into a centrifuge tube.
4. Lysis on ice for 30 min.
5. Centrifuge at 12000 x g for 5 min and collect the supernatant as total protein solution.

- **For suspension cell samples**

1. Centrifuge.to collect cells.
2. Mix the cytosol with RIPA Lysis Buffer (Strong) at a ratio of 250 μL of lysate per cell well of a 6-well plate and shake.
3. Ice bath for 30 min, repeatedly pipetting every 10 min to ensure complete cell lysis.
4. Centrifuge at 12000 x g for 5 min and collect the supernatant as total protein solution.

- **For bacterial or fungal samples**

1. Remove 1 mL of the suspension, centrifuge to remove the supernatant and wash once with PBS to fully remove the liquid. Vortex to disperse the bacterium as much as possible.
2. Add 100-200 μL of RIPA Lysis Buffer (Strong) and vortex gently to mix well with the lysate.
3. Ice bath for 30 min, repeatedly pipetting every 2 min to ensure complete cell lysis.
4. Centrifuge at 12000 x g for 5 min and collect the supernatant as total protein solution.

Note

1. Tissue or cells may appear viscous when lysed. This can be achieved by repeatedly blowing with a pipettor or shaking with a vortexer until it becomes a liquid. Add a further appropriate amount of lysis solution if it remains thick.
2. This reagent does not contain protease inhibitors. Add your own protease inhibitors before use. G2006, G2007, G2008 and other related protease inhibitors of our company are recommended.
3. For your safety and health, please wear safety glasses, gloves, or protective clothing.

По вопросам продаж и поддержки обращайтесь:

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Санкт-Петербург (812)309-46-40
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Саранск (8342)22-96-24
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Смоленск (4812)29-41-54
Сочи (862)225-72-31
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Сургут (3462)77-98-35
Сыктывкар (8212)25-95-17
Тамбов (4752)50-40-97
Тверь (4822)63-31-35
Тольятти (8482)63-91-07
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